



# Functional and Biochemical Endothelial Profiling *In Vivo* in a Murine Model of Endothelial Dysfunction; Comparison of Effects of 1-Methylnicotinamide and Angiotensin-converting Enzyme Inhibitor

Anna Bar<sup>1,2</sup>, Mariola Olkowicz<sup>3,4</sup>, Urszula Tyrankiewicz<sup>1</sup>, Edyta Kus<sup>1</sup>, Krzysztof Jasinski<sup>5</sup>, Ryszard T. Smolenski<sup>3</sup>, Tomasz Skorka<sup>5</sup> and Stefan Chlopicki<sup>1,2\*</sup>

## OPEN ACCESS

### Edited by:

Xinkang Wang,  
Agenix, USA

### Reviewed by:

Beat M. Jucker,

GlaxoSmithKline, England

Xiaoqiang Yao,

The Chinese University of Hong Kong,

Hong Kong

Andreas Daiber,

Johannes Gutenberg-Universität

Mainz, Germany

### \*Correspondence:

Stefan Chlopicki  
stefan.chlopicki@jcet.eu

### Specialty section:

This article was submitted to  
Cardiovascular and Smooth Muscle  
Pharmacology,  
a section of the journal  
Frontiers in Pharmacology

**Received:** 28 January 2017

**Accepted:** 21 March 2017

**Published:** 10 April 2017

### Citation:

Bar A, Olkowicz M, Tyrankiewicz U,  
Kus E, Jasinski K, Smolenski RT,  
Skorka T and Chlopicki S (2017)  
Functional and Biochemical  
Endothelial Profiling *In Vivo* in a  
Murine Model of Endothelial  
Dysfunction; Comparison of Effects  
of 1-Methylnicotinamide  
and Angiotensin-converting Enzyme  
Inhibitor. *Front. Pharmacol.* 8:183.  
doi: 10.3389/fphar.2017.00183

<sup>1</sup> Jagiellonian Centre for Experimental Therapeutics, Jagiellonian University, Krakow, Poland, <sup>2</sup> Chair of Pharmacology, Jagiellonian University Medical College, Krakow, Poland, <sup>3</sup> Department of Biochemistry, Medical University of Gdansk, Gdansk, Poland, <sup>4</sup> Department of Biotechnology and Food Microbiology, Poznan University of Life Sciences, Poznan, Poland, <sup>5</sup> Department of Magnetic Resonance Imaging, Institute of Nuclear Physics Polish Academy of Sciences, Krakow, Poland

Although it is known that 1-methylnicotinamide (MNA) displays vasoprotective activity in mice, as yet the effect of MNA on endothelial function has not been demonstrated *in vivo*. Here, using magnetic resonance imaging (MRI) we profile the effects of MNA on endothelial phenotype in mice with atherosclerosis (ApoE/LDLR<sup>-/-</sup>) *in vivo*, in comparison to angiotensin (Ang)-converting enzyme (ACE) inhibitor (perindopril), with known vasoprotective activity. On a biochemical level, we analyzed whether MNA- or perindopril-induced improvement in endothelial function results in changes in ACE/Ang II-ACE2/Ang-(1–7) balance, and L-arginine/asymmetric dimethylarginine (ADMA) ratio. Endothelial function and permeability were evaluated in the brachiocephalic artery (BCA) in 4-month-old ApoE/LDLR<sup>-/-</sup> mice that were non-treated or treated for 1 month or 2 months with either MNA (100 mg/kg/day) or perindopril (10 mg/kg/day). The 3D IntraGate®FLASH sequence was used for evaluation of BCA volume changes following acetylcholine (ACh) administration, and for relaxation time (T<sub>1</sub>) mapping around BCA to assess endothelial permeability using an intravascular contrast agent. Activity of ACE/Ang II and ACE2/Ang-(1–7) pathways as well as metabolites of L-arginine/ADMA pathway were measured using liquid chromatography/mass spectrometry-based methods. In non-treated 6-month-old ApoE/LDLR<sup>-/-</sup> mice, ACh induced a vasoconstriction in BCA that amounted to –7.2%. 2-month treatment with either MNA or perindopril resulted in the reversal of impaired ACh-induced response to vasodilatation (4.5 and 5.5%, respectively) and a decrease in endothelial permeability (by about 60% for MNA-, as well as perindopril-treated mice). Improvement of endothelial function by MNA and perindopril was in both cases associated with the activation of ACE2/Ang-(1–7) and the inhibition of ACE/Ang II axes as evidenced by an approximately twofold increase in Ang-(1–9) and Ang-(1–7) and a proportional decrease in Ang II and

its active metabolites. Finally, MNA and perindopril treatment resulted in an increase in L-arginine/ADMA ratio by 107% (MNA) and 140% (perindopril), as compared to non-treated mice. Functional and biochemical endothelial profiling in ApoE/LDLR<sup>-/-</sup> mice *in vivo* revealed that 2-month treatment with MNA (100 mg/kg/day) displayed a similar profile of vasoprotective effect as 2-month treatment with perindopril (10 mg/kg/day): i.e., the improvement in endothelial function that was associated with the beneficial changes in ACE/Ang II-ACE2/Ang (1–7) balance and in L-arginine/ADMA ratio in plasma.

**Keywords:** endothelial function, atherosclerosis, MRI, 1-methylnicotinamide, perindopril, plasma angiotensin profile, L-Arg/ADMA ratio

## INTRODUCTION

1-Methylnicotinamide (MNA), the major metabolite of NA synthesized in the liver in the reaction involving NNMT, when given exogenously has distinct therapeutic activity, despite the fact, that it has long been considered inactive (Aksoy et al., 1994). Indeed, topical MNA alleviates the inflammatory responses in skin diseases such as acne, contact dermatitis or rosacea (Gebicki et al., 2003; Wozniacka et al., 2005). In turn, systemic administration of MNA exerts anti-thrombotic (Chlopicki et al., 2007), anti-inflammatory (Bryniarski et al., 2008), and gastroprotective (Brzozowski et al., 2008) properties mediated by the activation of COX-2 and PGI<sub>2</sub> pathways. Furthermore, MNA can improve endurance exercise capacity in mice with diabetes, and may decrease the cardiovascular risk of exercise (Przyborowski et al., 2015). MNA was demonstrated to have hepatoprotective activity against concanavalin A-induced liver injury through the downregulation of IL-4 and TNF- $\alpha$  signaling (Sternak et al., 2010; Jakubowski et al., 2016), and to inhibit metastasis formation in a murine model of metastatic mammary gland cancer (4T1) in BALB/c mice (Blażejczyk et al., 2016). Interestingly, 1-MNA has also been shown to restore endothelial function in diabetic hypertriglyceremic rats (Bartuś et al., 2008) analyzed *ex vivo* in isolated aorta, suggesting that the improvement in endothelial function may represent an important target of MNA activity and may explain therapeutic efficacy of MNA in various diseases including diabetes (Watała et al., 2009) and atherosclerosis (Mateuszuk et al., 2016). Indeed, in our recent study anti-atherosclerotic effects of MNA in ApoE/LDLR<sup>-/-</sup> mice, including the inhibition

of inflammatory burden in plaques, diminished systemic inflammation, diminished platelet activation that were associated with an improvement in PGI<sub>2</sub> and NO-dependent endothelial function. These results support the notion that pronounced effects of MNA on endothelial function could contribute to therapeutic efficacy of MNA (Mateuszuk et al., 2016). However, in none of the previous experimental studies of MNA was endothelial function assessed *in vivo*, but only *in ex vivo* vascular preparation (Bartuś et al., 2008; Mateuszuk et al., 2016).

Endothelial dysfunction, being a consequence of vascular homeostatic imbalance is a hallmark of various cardiovascular diseases including atherosclerosis (Anderson et al., 1995), hypertension (Dharmashankar and Widlansky, 2010), heart failure (Marti et al., 2012), as well as non-cardiovascular diseases such as cancer (Franses et al., 2013). Vascular dysfunction is associated with activation of pro-inflammatory signaling molecules including adhesion molecules, chemokines and cytokines (Karch et al., 2014; Steven et al., 2015). There is also evidence that ROS play an important role in vascular inflammation (Zhou et al., 2011) and tissue damage (Mittal et al., 2014). There are number of enzymatic sources of endothelial ROS, including eNOS uncoupling, converting beneficial NO synthase into a detrimental superoxide-producing enzyme (Karch et al., 2014; Schulz et al., 2014).

Currently, many biochemical assays and functional tests are used for the evaluation of impaired NO-dependent function, oxidant stress and pro-inflammatory phenotype of endothelial dysfunction in humans. Prognostic value of biochemical measurements of endothelial biomarkers is still not widely accepted (Walczak et al., 2015). However, endothelial function measurements based on the assessment of impairment of NO-dependent vasodilatation in the coronary and peripheral circulation have prognostic importance in predicting adverse cardiovascular events (Schächinger et al., 2000; Halcox et al., 2002). In particular, a non-invasive endothelial function assessment, based on monitoring the brachial artery diameter with a two-dimensional ultrasound, before and after artery occlusion is considered a gold-standard method (Frolov et al., 2015). MRI has also successfully been used for non-invasive measurement of FMD in humans (Lee et al., 2007; Canton et al., 2012; Redheuil, 2014; Teixido-Tura et al., 2014), but studies of endothelial functional phenotype in mice *in vivo* are associated with greater technical challenges (Botnar and Makowski, 2012). Given high spatio-temporal resolution of MRI, it seems to be the method of choice to assess endothelium-dependent response in

**Abbreviations:** ACE, angiotensin-converting enzyme; ACE-I, angiotensin-converting enzyme inhibitor; Ach, acetylcholine; ACN, acetonitrile; ADMA, asymmetric dimethylarginine; Ang, angiotensin; ApoE/LDLR<sup>-/-</sup>, atherosclerotic mice; Arg, arginine; BCA, brachiocephalic artery; CA, contrast agent; COX-2, cyclooxygenase 2; CS, calibration standards; DDAH, dimethylarginine dimethylaminohydrolase; ECs, endothelial cells; eNOS, endothelial nitric oxide synthase; FA, formic acid; Hcy, homocysteine; HFD, high-fat diet; IL-4, interleukin-4; IS, internal standard; LC-MS/MS, liquid chromatography-tandem mass spectrometry; Met, L-methionine; MNA, 1-methylnicotinamide; MRI, magnetic resonance imaging; MRM, multiple reaction monitoring mode; NA, nicotinamide; NFPA, nonafluoropentanoic acid; NNMT, nicotinamide N-methyltransferase; NO, nitric oxide; Npx50, number of pixels, for which T<sub>1</sub> had changed more than 50% after contrast agent administration; PGI<sub>2</sub>, prostacyclin; QC, quality control samples; RAS, renin-angiotensin system; ROI, region of interest; ROS, reactive oxygen species; RS, real samples; SDMA, symmetric dimethylarginine; VFA, Variable Flip Angle technique; VSMCs, vascular smooth muscle cells; T<sub>1</sub>, relaxation time; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ .

mice. Indeed, MRI-based tests could provide an excellent tool to gain better insight into endothelium-dependent mechanisms in experimental studies in mice (Gotschy et al., 2013; Herold et al., 2009). Phinikaridou et al. (2012) were the first to describe an MRI-based method for the noninvasive assessment of endothelial-dependent vasodilatation and permeability *in vivo* in mice, with the use of an albumin-binding magnetic resonance CA. In our recent studies, we developed a 3D MRI-based assessment of endothelium-dependent vasodilatation induced by acetylcholine (ACh) and endothelial permeability fully based on the use of retrospectively self-gated 3D gradient-echo sequence (Bar et al., 2016). The developed approach has been validated in two distinct murine models of endothelial dysfunction, including atherosclerotic (ApoE/LDLR<sup>-/-</sup> mice) and HFD-fed mice, showing that our approach allows for a reliable detection of the impairment of NO-dependent vasodilatation and changes in endothelial permeability.

Genetically modified mice without apolipoprotein E and receptor for low density lipoprotein (mouse apolipoprotein E-deficient and Low-Density Lipoprotein Receptor-Deficient Double Knockout, ApoE/LDLR<sup>-/-</sup>), constitute reliable mouse model of endothelial dysfunction linked to atherosclerosis. In this strain of mice, spontaneously atherosclerosis develops without administration of atherogenic diet (Kostogrys et al., 2012), and endothelial dysfunction precedes the atherosclerotic plaque development (Csányi et al., 2012), similarly as it occurs in humans. In turn, the early development of endothelial dysfunction in single ApoE<sup>-/-</sup> mice was not univocally accepted (Crauwels et al., 2003; Villeneuve et al., 2003).

Given that the assessment of the endothelial phenotype *in vivo* in a reliable mouse model of endothelial dysfunction is essential for the preclinical profiling of vasoprotective compounds, the aim of the present study was to take advantage of the 3D MRI-based method to profile effects of MNA on functional phenotype of endothelium in ApoE/LDLR<sup>-/-</sup> mice *in vivo*, in comparison to standard vasoprotective treatment such as ACE-I (perindopril), that is known to display significant vasoprotective effect in various experimental models including atherosclerosis (Chłopicki and Gryglewski, 2005). We also analyzed whether MNA- or perindopril-induced improvement in endothelial function is associated with biochemical changes in plasma in ACE/Ang-II-ACE2/Ang-(1-7) balance and L-Arg/ADMA ratio.

## MATERIALS AND METHODS

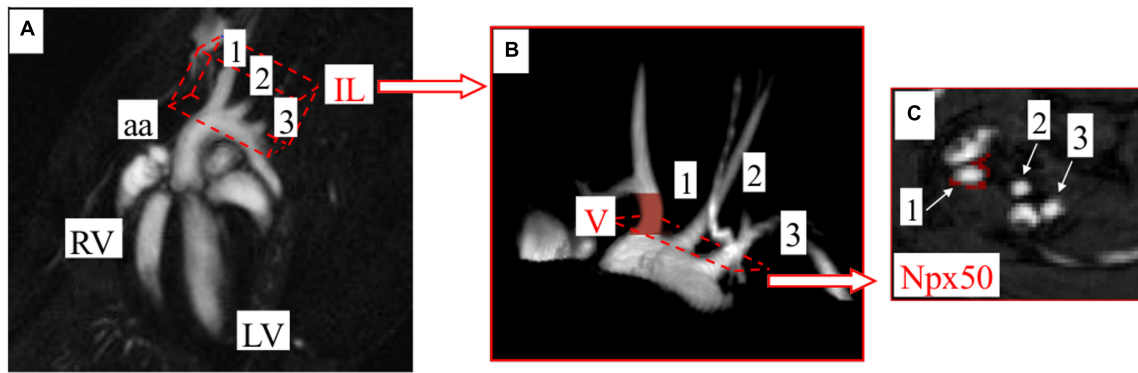
### Animals

In this study, 4-month-old ApoE/LDLR<sup>-/-</sup> mice (body weight of 20–30 g), bred in the Department of Human Nutrition, University of Agriculture (Krakow, Poland), were transported to the animal house at Jagiellonian Centre for Experimental Therapeutics (JCET), Jagiellonian University (Krakow, Poland) and were treated for 1 or 2 months with MNA (synthesized by dr Jan Adamus, Technical University in Lodz, Poland: 100 mg/kg/day in diet) or perindopril (a gift from Servier, Warszawa Poland: 10 mg/kg/day in drinking water). Doses of

compounds used in this study were based on our previous published (Chłopicki et al., 2007; Mateuszuk et al., 2016) and unpublished studies, for MNA and perindopril, respectively. Two groups of 4-month-old ApoE/LDLR<sup>-/-</sup> mice were left untreated (as control groups) for 1 and 2 months, respectively. The sizes and names of the experimental groups are reported in the legends of the corresponding graphs. All mice were bred in standard conditions (LD: 12/12, humidity: 60%, temperature: 23°C), and housed in pathogen-free conditions. Prior to the MRI experiment, the animals were transported to the animal house at the Institute of Nuclear Physics (Krakow, Poland) to assess endothelial phenotype *in vivo*. After *in vivo* measurements, mice were sacrificed in order to collect blood. The experimental design is illustrated in Supplementary Figure S1. All experiments were approved by the Ethics Local Committee of Jagiellonian University (Krakow, Poland) and were compliant with the Guide for the Care and Use of Laboratory Animals of the National Academy of Sciences (NIH publication No. 85–23, revised 1996).

### *In Vivo* Assessment of NO-dependent Endothelial Response to ACh and Endothelial Permeability by MRI

MRI experiments were performed using a 9.4T scanner (BioSpec 94/20 USR, Bruker, BioSpin GmbH, Germany), as described previously (Bar et al., 2016). Mice were anaesthetized using isoflurane (Aerrane, Baxter Sp. z o. o., Warszawa, Poland, 1.7 vol. %) in an oxygen and air (1:2) mixture. Body temperature was maintained at 37°C using circulating warm water. ECG, respiration and body temperature were monitored using a Model 1025 Monitoring and Gating System (SA Inc., Stony Brook, NY, USA). Mice were imaged in the supine position to test the vasomotor response of the vessel. 3D images of the aortic arch prior to and 25 min after intraperitoneal ACh administration (Sigma-Aldrich, Poznan Poland: 50 µl, 16.6 mg/kg) were acquired using the cine IntraGate<sup>TM</sup> FLASH 3D sequence and reconstructed with the IntraGate 1.2.b.2 macro (Bruker, BioSpin GmbH, Germany). T<sub>1</sub> maps were measured before and 30 min after intravenous administration of albumin-binding gadolinium CA (Galbumin, BioPal, Worcester, MA, USA – 25 mg/ml, 4.5 ml/kg) using the VFA technique (Cheng and Wright, 2006; Wang et al., 2006), by sampling the signal, using varying values of flip angles and then fitting the result to an expected T<sub>1</sub>-dependent signal model. Imaging parameters for endothelial function assessment included the following: repetition time (TR) – 6.4 ms, echo time (TE) – 1.4 ms, field of view (FOV) – 30 mm × 30 mm × 5 mm, matrix size – 256 × 256 × 30, flip angle (FA) – 30°, and number of accumulations (NA) – 15, reconstructed to seven cardiac frames. Total scan time was 10 min. Imaging parameters for endothelial permeability assessment included the following: TR – 10 ms, TE – 1.1 ms, FOV – 30 mm × 30 mm × 4 mm, matrix size – 192 × 160 × 8, number of repetitions – 12, and reconstructed to one cardiac frame. Eight FA were used: 2°, 4°, 6°, 8°, 14°, 20°, 30°, 50°. FA values were set by changing the length of a radiofrequency pulse, with constant amplifier power. Total scan time for all angles was



**FIGURE 1 | Methodology to assess endothelium-dependent response and changes in endothelial permeability in brachiocephalic artery (BCA) *in vivo* by MRI. (A)** Coronal view of the heart. Image showing position of the imaging layer (IL) used for 3D imaging of the aortic arch. **(B)** 3D image of aortic arch acquired with the cine IntraGate® FLASH 3D sequence. Endothelial function assessment, expressed as changes in the vessels volume (V) can be performed in vessels arising from aortic arch. The end-diastolic volumes of BCA were assessed prior to and 25 min after intraperitoneal Ach administration. **(C)** Representative images of vessels cross-sections, in which a number of pixels, for which  $T_1$  had changed more than 50% after CA administration (Npx50), is marked in red. BCA is indicated as 1, left carotid artery (LCA) as 2, left subclavian artery (LSA) as 3. RV, right ventricle; LV, left ventricle.

16 min. The experimental design is illustrated in Supplementary Figure S1.

## MRI Data Analysis

Time-resolved 3D images of the aortic arch (**Figure 1A**) were analyzed to endothelial function assessment in BCA, using ImageJ software 1.46r (NIH Bethesda, MD, USA) and scripts written in Matlab (MathWorks, Natick, MA, USA). Images were reconstructed to seven cardiac frames and imported into ImageJ as a hyperstack (**Figure 1B** matrix:  $256 \times 256$ , slices: 30, frames: 7). Further analysis was performed in diastole of BCA, using small hyperstack (**Figure 1B** marked in red color matrix:  $256 \times 256$ , slices: 5, frames: 1) starting at the base of the vessel and ending just before the branch. Cross-sectional areas of BCA at each slices were obtained using thresholding segmentation and exported to Matlab, where BCA volumes were reconstructed and calculated. Detailed analysis was described in Supplementary Material of our previous work (Bar et al., 2016). In order to endothelial permeability assessment, obtained images were used to calculate the  $T_1$  around the BCA lumen before and after CA administration. The signal model was fitted pixel by pixel using Matlab software developed in house. Two  $T_1$  maps (before and after CA administration) were compared, using scripts written in Matlab, and pixels for which  $T_1$  had changed significantly (by more than 50%) after CA administration were marked in red (**Figure 1C**). The threshold value (50%) was determined experimentally. All red pixels were counted by program as number of pixels, for which  $T_1$  had changed more than 50% after CA administration (Npx50).

## Measurement of Ang Profile in Plasma

The blood was drawn from the heart, collected in tubes containing heparin (20 units/ml) and immediately mixed with protease inhibitor cocktail (Sigma-Aldrich, Poznan, Poland) in a ratio of 19:1 (v/v), and centrifuged at  $1,000 \times g$  for

10 min to isolate plasma. Afterward, the resulting plasma was transferred into Protein LoBind tubes (Eppendorf, Hamburg, Germany), split into aliquots and stored frozen at  $-80^\circ\text{C}$  until analysis.

Protein precipitation with ACN was used as the sample pre-treatment method to remove high molecular plasma compounds. CS, QC samples, and RS were created by spiking 25  $\mu\text{l}$  (pooled blank – CS, QC) plasma with 20  $\mu\text{l}$  appropriate working standard solution or water (RS) and 5  $\mu\text{l}$  IS – [Asn<sup>1</sup>, Val<sup>5</sup>]-Ang II (2,500 pg/ml). Calibration curve standards were made at: 10; 50; 100; 250; 300; 400; 500 pg/ml (IS – 250 pg/ml) concentrations, respectively. CS at seven different concentration levels or adequate QC and RS were deproteinized with ACN (in 4:1 (v/v) proportion to the samples used) and assayed.

The LC-MS/MS analyses of plasma samples were performed on an UltiMate 3000 Rapid Separation nano-LC system (Dionex, Thermo Scientific, San Jose, CA, USA) interfaced via a ChipMate™ nanoelectrospray ion source (Advion, USA) to a TSQ Vantage triple quadrupole mass spectrometer (Thermo Scientific). The samples were injected in 5  $\mu\text{l}$  aliquots onto a trapping column for desalting and concentrating of the analytes which in the next step was switched in-line with the separation C18 column to elute the peptides at a flow rate of 300 nl/min with an increasing percentage of the organic solvent. Separation was accomplished using a gradient of phase A (acetic acid (1%, v/v) in H<sub>2</sub>O) and phase B (acetic acid (1%, v/v) in ACN) as follows: 2% B for 5 min, 2–98% B for 15 min, and 98% B for 5 min. The mass spectrometer was operated in the positive ion mode and the detection of Ang peptides in MRM mode was performed. The two most sensitive/specific ion transitions were measured for each of the 9 Ang peptides determined [Ang I, II, III, IV, (1–7), (1–9), Ang A, alamandine and Ang-(1–12)]. Analytical method details as well as the MS/MS transitions monitored in the protocol are described elsewhere (Olkowicz et al., 2016). Instrument control, data collection, and analysis were achieved with the Thermo Xcalibur (version 2.1) software.



## Measurement of Flux Through L-Arg Metabolic Pathways

Plasma amino acid concentrations were determined using a Surveyor HPLC system (Thermo Scientific) coupled to a TSQ Vantage Triple-Stage Quadrupole mass spectrometer, as described recently (Smolenska et al., 2016). Briefly, an aliquot of plasma (50  $\mu$ l) was deproteinized with 100  $\mu$ l of ACN maintained in ice. The tubes were then centrifuged at 4°C, 16,000  $\times$  g for 10 min and the resultant supernatant was collected and subjected to freeze-drying. The residue was next reconstituted with 50  $\mu$ l of H<sub>2</sub>O acidified with 0.1% (v/v) FA and analyzed using an LC/MS system as detailed below.

Chromatographic separation was achieved using a Synergi Hydro-RP column (50 mm  $\times$  2.0 mm i.d., 2.5  $\mu$ m) fitted with a security guard (Phenomenex, Torrance, CA, USA). The mobile phase consisted of a mixture of phase A (5 mM NFPA in water) and phase B (0.1% (v/v) FA in ACN), that was delivered at 0.2 ml/min in a gradient from 0 to 45% B in 5.5 min. The effluent of the HPLC column was directed to the electrospray Ion-Max source of the mass spectrometer that was operated in MRM mode. The settings of the ion source were as follows: needle voltage, +4.5 kV; sheath gas pressure, 30 (arbitrary units); auxiliary gas pressure, 3 (arbitrary units), and capillary temperature, 250°C. Additionally, to overcome the possible reduction of the signal intensity due to perfluorinated carboxylic acid use, we added 0.05% (v/v) FA in methanol at 0.2 ml/min as the post-column sheath flow. The LC/MS system, data acquisition and processing were managed by the Xcalibur software (v. 2.1, Thermo Scientific).

## Statistical Analysis

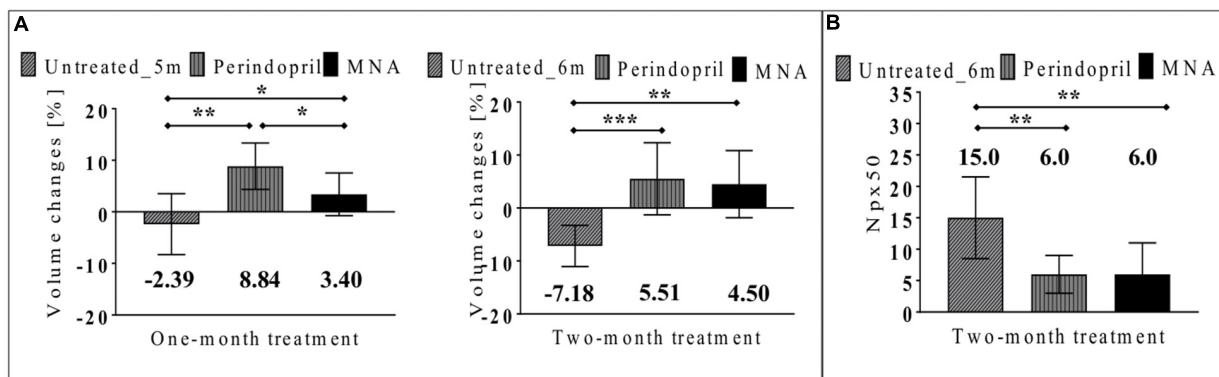
All of the data obtained are presented as mean and standard error of the mean (SEM) or in case of the lack of normal distribution as median and range. Statistical tests were

done using STATISTICA 10 (Stat Soft Inc., USA) or GraphPad Prism 5 (GraphPad Software, Inc., CA, USA) software. Non-parametric test (Kruskal–Wallis test) or parametric test (one-way ANOVA followed by Tukey's *post hoc* test) were performed. Pearson or Spearman correlation coefficient tests were used to assess dependence between two parameters. Statistical significance was defined as  $p < 0.05$ .

## RESULTS

### Effects of MNA and Perindopril Treatment on Endothelium-dependent Vasomotor Response and Changes in Endothelial Permeability in ApoE/LDLR<sup>-/-</sup> Mice *In Vivo*

In non-treated ApoE/LDLR<sup>-/-</sup> mice, injection of Ach (16.6 mg/kg given i.p. in the volume of 50  $\mu$ l) resulted in vasoconstriction of BCA. Endothelium-dependent response of BCA after Ach administration amounted to -2.39 and -7.18% for 5-month-old and 6-month-old ApoE/LDLR<sup>-/-</sup> mice, respectively. In mice treated with MNA or perindopril for a 1-month period (Figure 2A), endothelium-dependent response improved and an increase in volume of BCA after Ach injection was observed (by about 9 and 3%, respectively). Treatment of ApoE/LDLR<sup>-/-</sup> mice for 2 months with MNA or perindopril further improved endothelium-dependent vasodilation as well as decreased endothelial permeability (Figure 2B). The number of pixels around BCA, for which T<sub>1</sub> had changed more than 50% after CA administration (Npx50) was smaller in mice treated with MNA and perindopril than in non-treated ApoE/LDLR<sup>-/-</sup> mice (Npx50 was 6.0, 6.0, and 15.0, respectively, Figure 2B). Interestingly, there was a significant correlation between endothelium-dependent response of BCA and Npx50 (Table 1).



**FIGURE 2 | Effects of MNA and ACE-I (perindopril) treatment on endothelial function and permeability *in vivo* in ApoE/LDLR<sup>-/-</sup> mice.** Changes in end-diastolic volume of BCA 25 min after Ach administration (A) and number of pixels around BCA, for which T<sub>1</sub> had changed more than 50% after CA administration (Npx50) (B) in non-treated ApoE/LDLR<sup>-/-</sup> mice (columns with diagonal lines) and in ApoE/LDLR<sup>-/-</sup> mice treated with MNA (black columns) or perindopril (columns with vertical lines) for one month (Untreated\_5m  $n = 8$ , perindopril  $n = 5$ , MNA  $n = 6$ ) or two months (Untreated\_6m  $n = 8$ , perindopril  $n = 9$ , MNA  $n = 7$ ). Statistics: Kruskal–Wallis test, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

**TABLE 1 | Correlation between parameters of functional and biochemical endothelial profiling *in vivo*.**

The pair of parameters	R Spearman	p
BCA volume changes and Npx50	-0,668739	0,000668
BCA volume changes and L-arginine/ADMA ratio	0,765473	0,000000
BCA volume changes and Ang-(1-7)/Ang II ratio	0,710365	0,000001
Npx50 and L-arginine/ADMA ratio	-0,770589	0,000070
Npx50 and Ang-(1-7)/Ang II ratio	-0,639038	0,002420

## Effects of MNA and Perindopril Treatment on Plasma Ang Profile

The pattern of changes in plasma Ang profile of ApoE/LDLR<sup>-/-</sup> mice treated for one month with MNA was largely convergent with those observed in plasma of mice treated with perindopril (as compared to non-treated animals), and involved an increase in concentration of Ang-(1-7) [by 36% (MNA), 95% (perindopril)], alamandine [by 18% (MNA), 47% (perindopril)], and Ang-(1-9) [by 72% (MNA), 173% (perindopril)] in comparison to untreated ApoE/LDLR<sup>-/-</sup> mice (**Figure 3A**). At the same time, the level of Ang II and other pathological effectors of the RAS (Ang III, IV, A) significantly decreased. The most pronounced differences were observed in the content of Ang II and its N-terminally truncated metabolite – Ang III, the level of which nearly halved as compared to the control, untreated animals [from 119.8 ± 11.3 to 85.5 ± 5.4 (MNA) and to 50.2 ± 4.1 (perindopril) fmol/ml for Ang II, from 206.2 ± 15.5 to 130.4 ± 10.0 (MNA), and to 96.2 ± 8.7 (perindopril) fmol/ml for Ang III, respectively]. These changes were accompanied by a slight increase in the content of precursor peptides: Ang I [by 21% (MNA) and 34% (perindopril) versus non-treated mice] and Ang-(1-12) [by 18% (MNA) and 43% (perindopril) as compared to control animals]. Activation of counter-regulatory ACE2/Ang-(1-7)/Ang-(1-9) axis with concomitant inhibition of ACE/Ang II pathway was further highlighted after two months of treatment with MNA or perindopril whereby the beneficial effect of MNA treatment was even more similar to that exerted by perindopril (**Figure 3B**). Interestingly, there was a significant correlation between Ang-(1-7)/Ang II ratio and endothelium-dependent response of BCA or Npx50 (**Table 1**).

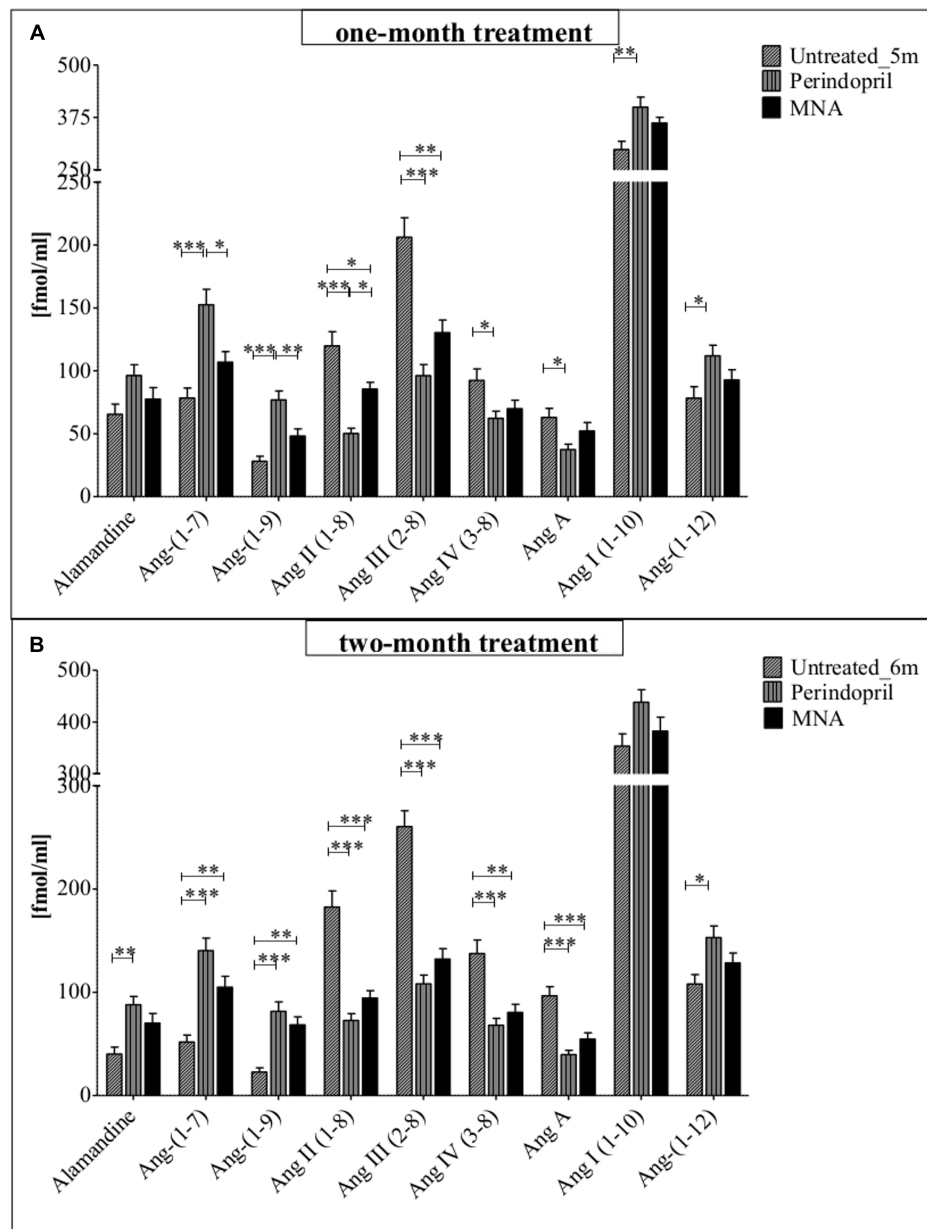
## Effects of MNA and Perindopril Treatment on L-Arg/ADMA Ratio in Plasma

Endothelial function is critically dependent on eNOS activity and consequently adequate NO bioavailability. Treatment with both MNA and perindopril was associated with alterations in amino acid metabolism engaged in NO generation. Plasma concentration of L-Arg that serves as a substrate for the formation of NO by the NOS enzymes was elevated after just one month of treatment with both MNA (by 25%) and perindopril (21%) as compared to untreated ApoE/LDLR<sup>-/-</sup> mice (**Figure 4A**). Furthermore, the plasma concentration of precursors of L-Arg (L-citrulline and L-ornithine) was increased in parallel. These changes were accompanied by a decrease

in methylated Arg concentration. After 1 month of treatment, ADMA that is perceived as an endogenous inhibitor of all NOS enzymes (including eNOS) was diminished by 10% (MNA) and 17% (perindopril), respectively (in comparison to non-treated animals). Two months of pharmacotherapy led to a further decline in the plasma concentration of ADMA [by 15% (MNA) and by 20% (perindopril), respectively] (**Figure 4B**). Plasma concentration of SDMA, recognized as a competitive inhibitor of cellular L-Arg transport was also decreased after 1 and 2 months of treatment with MNA or perindopril, but these changes did not reach statistical significance. Moreover, reduced concentration of methylated Arg was associated with an increase in the content of Met and concomitant decrease in the level of Hcy. Consequently, plasma L-Arg/ADMA ratio reflecting NO bioavailability was clearly elevated after 1 month of treatment with both MNA (by 34%) and perindopril (by 46%), and this effect was further maintained after 2 months of treatment. Interestingly, there was a significant correlation between L-arginine/ADMA ratio and endothelium-dependent response of BCA or Npx50 (**Table 1**).

## DISCUSSION

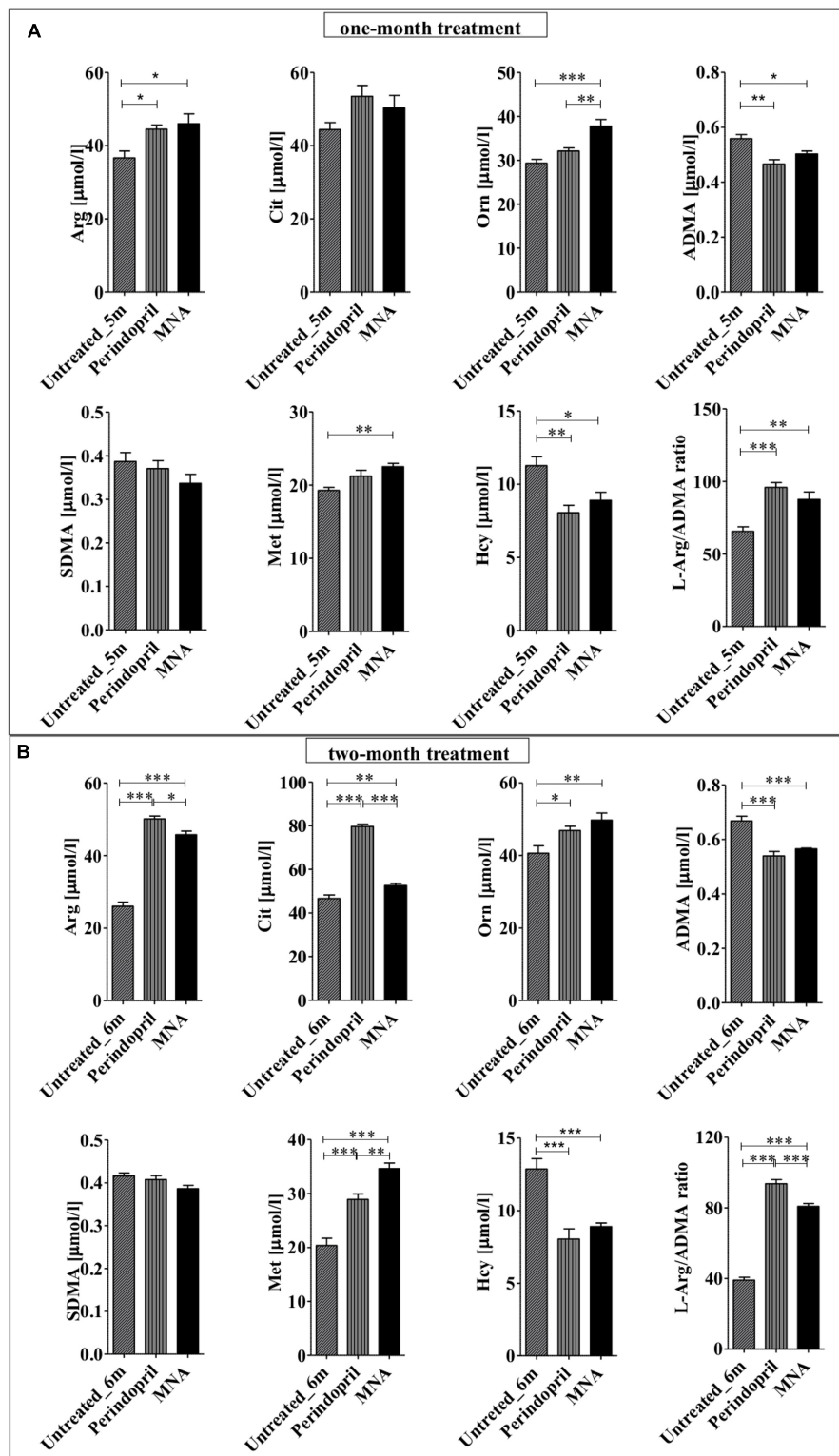
In the present work, we profiled endothelial activity of MNA in comparison with perindopril (ACE-I) using functional 3D MRI-based *in vivo* assays of endothelial function in parallel to biochemical measurements of ACE/Ang II-ACE2/Ang-(1-7) balance, and L-Arg/ADMA ratio in plasma. We demonstrated *in vivo* in ApoE/LDLR<sup>-/-</sup> mice, that treatment with MNA or perindopril resulted in the reversal of Ach-induced vasoconstriction response to vasodilatation and a concomitant decrease in endothelial permeability. Functional improvement of endothelial function by MNA or perindopril treatment was in both cases associated with the inhibition of ACE/Ang II axis and parallel activation of ACE2/Ang-(1-7) axis as evidenced by a fall in plasma concentration of Ang II, and its N-terminally truncated metabolites (Ang III and IV) and an increase in Ang-(1-9) and Ang-(1-7) plasma concentration. Furthermore, MNA and perindopril treatment resulted in an increase in L-arginine/ADMA ratio compatible with improvement of NO-dependent endothelial function. Altogether, our results suggest that MNA displayed a similar pattern of vasoprotective activity as perindopril, a representative of ACE-Is, and both compounds affected ACE/Ang II-ACE2/Ang-(1-7) balance and L-Arg/ADMA ratio that could contribute to their vasoprotective effects. Given that correction of endothelial function by ACE-Is contributes significantly to their clinical benefits and their anti-inflammatory, anti-thrombotic, anti-diabetic, and vasoprotective actions reported in numerous experimental and clinical papers (Chłopicki and Gryglewski, 2005), the findings of endothelial profile of MNA being similar to perindopril in *in vivo* settings bring important novel perspectives to understanding the therapeutic efficacy of MNA reported previously (Gebicki et al., 2003; Wozniacka et al., 2005; Chłopicki et al., 2007; Bartuś et al., 2008; Bryniarski et al., 2008; Brzozowski et al., 2008; Watała et al., 2009; Sternak et al., 2010; Przyborowski et al., 2015; Błazejczyk et al., 2016; Jakubowski et al., 2016; Mateuszuk et al., 2016).



**FIGURE 3 | Effects of MNA and ACE-I (perindopril) treatment on plasma Ang profile in ApoE/LDLR<sup>-/-</sup> mice.** Plasma Ang profile in non-treated ApoE/LDLR<sup>-/-</sup> mice (columns with diagonal lines) and in ApoE/LDLR<sup>-/-</sup> mice treated with MNA (black columns) or perindopril (columns with vertical lines) for one month (**A**: Untreated\_5m  $n = 6$ , perindopril  $n = 5$ , MNA  $n = 5$ ) or two months (**B**: Untreated\_6m  $n = 6$ , perindopril  $n = 8$ , MNA  $n = 7$ ). Statistics: one-way ANOVA followed by Tukey's *post hoc* test (normality was assessed using the Kolmogorov-Smirnov test); \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .

In this study, endothelial function *in vivo* assessment was performed in BCA, in response to intraperitoneal administration of Ach. Importantly, endothelium-dependent response was independent of the effect of Ach on the heart and respiration (Bar et al., 2016). Undoubtedly, the important advantage of the use of MRI-based method is a retrospective reconstruction of images from 3D data sets, which allows for assessment of response of the entire vessel instead of only one fragment of the vessel. Moreover, the method provides the possibility of

accurate positioning after experiment, whereby the effects of endothelial pharmacotherapy can be assessed, even in small vessels such as BCA, across various cross-sections to be located in planes perpendicular to the vessel. The 3D gradient echo sequence allows the avoidance of precise slice positioning during measurements, as arbitrary re-slicing during post-processing is possible with shorter time of the measurements. Importantly, in the present work an MRI-based method to assess endothelial permeability changes was also employed. We assessed the



**FIGURE 4 | Effects of MNA and ACE-I (perindopril) treatment on L-arginine/ADMA ratio in plasma in ApoE/LDLR<sup>-/-</sup> mice.** Plasma concentration of L-Arg, its precursors (L-Cit, L-Orn), ADMA, SDMA, Met, Hcy as well as L-Arg/ADMA ratio in non-treated ApoE/LDLR<sup>-/-</sup> mice (columns with diagonal lines) and in ApoE/LDLR<sup>-/-</sup> mice treated with MNA (black columns) or perindopril (columns with vertical lines) for one month (**A**: Untreated\_5m  $n = 6$ , perindopril  $n = 5$ , MNA  $n = 5$ ) or two months (**B**: Untreated\_6m  $n = 6$ , perindopril  $n = 8$ , MNA  $n = 7$ ). Statistics: one-way ANOVA followed by Tukey's *post hoc* test (normality was assessed using the Kolmogorov-Smirnov test); \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ .



number of pixels (around the BCA) for which  $T_1$  had changed more than 50% (Npx50), as described previously (Bar et al., 2016). Npx50 parameter allows avoiding precise determination of ROI, what is difficult to establish objectively. Finding the pixels for which  $T_1$  had changed more than 50%, allows for operator-independent assessment of the ROI around the vessel. Analysis of changes in endothelial permeability represents an important feature of endothelial dysfunction, and its measurements seem a valuable test to analyze endothelial state *in vivo*. When endothelial permeability increases, leaky intercellular junctions of endothelium allow access to low-density lipoprotein particles and transmigration of leukocytes that are critical in atherosclerotic plaque formation (Kao et al., 1995). Moreover, albumin-binding CA can also accumulate in vessel walls (Lobbes et al., 2009; Pedersen et al., 2011), and this results in shortening of the  $T_1$  in the vessel wall enabling MRI-based detection, and Npx50-based operator-independent assessment of endothelial permeability (Bar et al., 2016). Importantly, in the present work, analysis of endothelial-dependent response by Ach as well as endothelial permeability based on Npx50 displayed significant negative correlation and gave concordant results showing improvement after MNA and perindopril treatment. Experiments carried out in this paper support the notion that MRI-based technique based on the retrospectively self-gated 3D IntraGate® FLASH sequence (Bar et al., 2016) is useful for monitoring the efficacy of endothelium-targeted therapy in murine models of diseases *in vivo* and seems far superior to ultrasound (Cavalcante et al., 2011).

In non-treated ApoE/LDLR<sup>-/-</sup> mice, injection of Ach resulted in paradoxical vasoconstriction, what was most likely due to a smooth muscle muscarinic receptor-dependent response, as also shown for human arteries with endothelial dysfunction (Ludmer et al., 1986). Treatment with MNA or perindopril resulted in shifting the Ach response to a vasodilatation, which indicates improvement in endothelial function. Comparing results of changes in BCA volume after acetylcholine administration in 5-month-old C57BL/6J mice (9.3%) published in our previous paper (Bar et al., 2016), to ApoE/LDLR<sup>-/-</sup> mice after two-month treatment with MNA (4.5%) or perindopril (5.5%) performed in present study, it seems that either treatment considerably improved the impaired endothelium-dependent response. It is worth noting that, after one-month treatment effects of perindopril seemed to be more pronounced as compared with the effect of MNA, but this difference disappeared after two months of treatment. Improvement in endothelial permeability was also similar in magnitude for MNA and perindopril after two months of treatment, supporting the similar pattern of endothelial response to MNA given in a dose of 100 mg/kg b.w./day in comparison to perindopril given in a dose of 10 mg/kg b.w./day.

An important finding of the present study was the demonstration that improvement of endothelial function by MNA and perindopril treatment, evidenced by MRI-based assays, was in both cases associated with the inhibition of the classical RAS pathway – ACE/Ang II/AT<sub>1</sub>R and concomitant activation of counter-regulatory RAS pathway – ACE2/Ang-(1–7)/Mas. Numerous previous studies clearly indicate that the

RAS plays a critical role in the initiation and progression of atherosclerosis with endothelial dysfunction as its early event, and that the balance between two major axes of RAS, i.e., ACE/Ang II and ACE2/Ang-(1–7), is a key determinant of RAS beneficial vs. detrimental effects (Wang et al., 2013; Olkiewicz et al., 2015; Cahill and Redmond, 2016). Ang II is considered as a pro-atherosclerotic mediator which causes vasoconstriction, regulates adhesion molecule (ICAM-1, VCAM-1, P-selectin) expression as well as chemokine, cytokine, and growth factor secretion within the arterial wall (Jiang et al., 2014). On the other hand, ACE2, through its product Ang-(1–7) displays anti-atherosclerotic properties *in vivo*, including decreasing inflammation and oxidative stress, and inhibiting inflammatory cell infiltration. The protective effect of ACE2, mainly distributed in ECs, VSMCs, and macrophages, was repeatedly demonstrated. Overexpression of ACE2 by gene transfer attenuated the progression of atherosclerotic lesions when the ACE2 gene was transferred 4 weeks after injury in a rabbit model of atherosclerosis (Dong et al., 2008). Similarly, overexpression of ACE2 in ApoE-deficient mice decreased atherosclerotic lesion size within the aortic sinus as compared to control mice (Lovren et al., 2008). In contrast, ACE2-deficiency in LDLR<sup>-/-</sup> as well as ApoE<sup>-/-</sup> genetic background mice increased the development of atherosclerosis (Thomas et al., 2010; Thatcher et al., 2011).

Our results not only confirmed that perindopril-induced a vasoprotective shift in the balance of ACE/Ang II and ACE2/Ang-(1–7), but also underscored that the profile of MNA is similar to that exerted by perindopril. Zhuo et al. (2002) have recently demonstrated that the clinical benefits of chronic ACE inhibition with perindopril, in addition to the blockade of systemic and tissue Ang II formation, may be in part due to elevated bradykinin levels, which in turn lead to increased NO bioavailability. Bradykinin was shown to be the major mediator of ACE-I-induced anti-thrombotic effects (Gryglewski et al., 2003, 2005; Chłopicki and Gryglewski, 2005). We did not study here the relative contribution of Ang II-dependent and bradykinin-dependent pathway to perindopril-induced improvement in NO-dependent function, but obviously it cannot be excluded that both pathways may be involved in perindopril-induced improvement in endothelial function. Whether the same mechanisms contribute to MNA-induced effects remains to be established. Interestingly, in our previous studies describing anti-atherosclerotic effects of MNA, an improvement in NO-dependent endothelial function, observed in *ex vivo* vascular preparations was also associated with activation of PGI<sub>2</sub>-dependent function (Mateuszuk et al., 2016). Altogether, the results presented here suggest that MNA-induced shift in the balance of ACE/Ang II and ACE2/Ang-(1–7) toward the vasoprotective axis may contribute to the improvement of endothelial function by MNA or alternatively may constitute an important pathophysiological sign of improved endothelial function.

We also demonstrated that MNA and perindopril treatment resulted in an increase in L-arginine/ADMA ratio that was compatible with improvement of NO-dependent endothelial function. Given the fact, that circulating level of ADMA adds independent prognostic information with regard to

cardiovascular risk beyond that obtained from classical risk factors and novel biomarkers (Schnabel et al., 2005) this finding is of importance. In fact, plasma concentrations of ADMA have been found to be significantly elevated in patients with arteriosclerosis, as well as in those with coronary risk factors, such as hypertension or hypercholesterolemia, and marked increase in ADMA level has been associated with impaired endothelium-dependent NO-mediated vasodilatation (Böger et al., 1997, 1998; Surdacki et al., 1999; Böger, 2003) that may be linked to decreased activity of  $\gamma$ -LAT as the major export pathway for the ADMA (Closs et al., 2012). Interestingly, Suda et al. (2004) have recently reported that the long-term vascular effects of ADMA are not solely mediated by inhibition of endothelial NO synthesis but also can lead to direct upregulation of ACE expression and increased oxidative stress through AT<sub>1</sub> receptor activation. Moreover, they have proposed that inhibition of NO synthesis appears primarily as a result of short-term action of ADMA; as long as metabolites of ADMA accumulate at higher concentrations in blood vessels, eNOS-independent mechanisms predominate. Involvement of multiple mechanisms in vascular effects of ADMA, other than simple inhibition of endothelial NO synthesis, including also upregulation of ACE, remains to be clarified. In a recent work of Jiang et al. (2016) it was reported that MNA attenuated atherogenesis *via* modulation of the ADMA-DDAH pathway in ApoE-deficient mice. The beneficial effect of MNA treatment was associated with the upregulation of the DDAH2 enzyme activity in endothelium *via* the mitigation of hypermethylation in the promoter region of the enzyme. These results seem consistent with our observations showing the reduction in plasma ADMA concentration as well as Hcy concentration that was linked to an improved endothelium-dependent vascular relaxation after MNA. Interestingly, Huang et al. (2015) showed that Hcy synthesized in endothelium (after Met administration) affected activity of ACE by direct homocysteinylation of its amino- and/or sulfhydryl- moieties. This modification enhanced ACE reactivity toward Ang II and consequently led to NADPH oxidase-superoxide-dependent endothelial dysfunction. In that context, treatment with MNA and perindopril attenuates various apparently interlinked biochemical changes closely associated with functional phenotype of endothelial dysfunction such as excessive activation of ACE, elevation of ADMA and Hcy that can all contribute to vasoprotective action of these compounds resulting in the improvement of endothelial function reported here *in vivo* using functional and biochemical measurements.

Indeed, there was a significant positive correlation between endothelium-dependent response of BCA after acetylcholine administration and the two major biochemical parameters measured here [L-arginine/ADMA ratio and Ang-(1-7)/Ang II]. In turn, since the improvement of endothelial dysfunction was associated with a decrease in endothelial permeability, correlation between Npx50 and biochemical parameters [L-arginine/ADMA ratio and Ang-(1-7)/Ang II] had negative character. Similar correlations between ADMA concentration and Ach- or flow-induced vasodilatation have also been demonstrated previously in monkeys with hypercholesterolemia, using endothelium-dependent vascular function *ex vivo* assessment (Böger et al.,

2000) as well as in humans with hypercholesterolemia using ultrasonography (Böger et al., 1998).

Effects of treatment with MNA and perindopril on atherosclerotic plaque burden were not assessed in this study, but it was reported previously that perindopril treatment inhibited the development of atherosclerotic lesions in diabetic ApoE-deficient mice (Candido et al., 2002), while anti-atherosclerotic effect of MNA was described in apolipoprotein E (ApoE)/low-density lipoprotein receptor (LDLR)-deficient mice (Mateuszuk et al., 2016). Given the pathophysiological role of endothelial dysfunction in atherogenesis, effects of MNA or perindopril on endothelial function could contribute to the anti-atherosclerotic action of these compounds.

## CONCLUSION

Using functional 3D MRI-based assays of endothelial function and biochemical measurements of ACE/Ang II, ACE2/Ang-(1-7) balance and L-Arg/ADMA ratio in plasma, we demonstrated significant vasoprotective effects of MNA (100 mg/kg b.w./day), which were comparable with that afforded by perindopril (10 mg/kg b.w./day), a well-known ACE inhibitor. Although effects of MNA were less pronounced as compared with perindopril after 1-month treatment, after 2-month treatment, either treatment modality resulted in a comparable effect including reversal of Ach-induced vasoconstriction response to vasodilatation and a concomitant decrease in endothelial permeability. Furthermore, functional improvement of endothelial function by MNA or perindopril treatment was in both cases associated with the inhibition of ACE/Ang II axis, the parallel activation of ACE2/Ang-(1-7) axis, and an increase in L-Arg/ADMA ratio in plasma. Given that improvement in endothelial function by ACE-Is contributes to their therapeutic efficacy (Chłopicki and Gryglewski, 2005), presented in the current study findings of endothelial profile of MNA being similar to ACE-I (perindopril) in *in vivo* settings bring important novel perspectives to understand therapeutic efficacy of MNA (Gebicki et al., 2003; Wozniacka et al., 2005; Chłopicki et al., 2007; Bartuś et al., 2008; Bryniarski et al., 2008; Brzozowski et al., 2008; Watała et al., 2009; Sternak et al., 2010; Przyborowski et al., 2015; Błazejczyk et al., 2016; Jakubowski et al., 2016; Mateuszuk et al., 2016). Moreover, our approach for the *in vivo* assessment of endothelial phenotype based on a retrospectively self-gated 3D gradient-echo sequence, MRI-based technique, concomitant with the biochemical assays of two important systems of endothelial regulation, ACE/Ang-II-ACE2/Ang-(1-7) and L-Arg/ADMA, proves to be a useful tool for the *in vivo* endothelial profiling of compounds to demonstrate convincingly their beneficial effects on endothelial function.

## AUTHOR CONTRIBUTIONS

Conceived and designed the study: AB and SC. Performed the study: AB, MO, UT, EK, and KJ. Analyzed the data: AB and MO. Provided the analytical tools: RS, TS. Drafted the manuscript: AB,

MO, and SC. All authors have corrected or have approved the final version of the manuscript.

## ACKNOWLEDGMENTS

This study was supported by the National Science Centre, grant Symfonia no. DEC-2015/16/W/NZ4/00070 and partially by the European Union from the resources of the European Regional Development Fund under the Innovative Economy Programme

(grant coordinated by JCET-UJ, No POIG. 01.01.02-00-069/09). We would like to thank Renata Kostogrys for support and assistance.

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fphar.2017.00183/full#supplementary-material>

## REFERENCES

- Aksoy, S., Szumlanski, C. L., and Weinshilboum, R. M. (1994). Human liver nicotinamide N-methyltransferase. cDNA cloning, expression, and biochemical characterization. *J. Biol. Chem.* 269, 14835–14840.
- Anderson, T. J., Gerhard, M. D., Meredith, I. T., Charbonneau, F., Delagrang, D., Creager, M. A., et al. (1995). Systemic nature of endothelial dysfunction in atherosclerosis. *Am. J. Cardiol.* 75, 71B–74B. doi: 10.1016/0002-9149(95)80017-M
- Bar, A., Skorka, T., Jasinski, K., Sternak, M., Bartel, Ż., Tyrankiewicz, U., et al. (2016). Retrospectively-gated MRI for in vivo assessment of endothelium-dependent vasodilatation and endothelial permeability in murine models of endothelial dysfunction. *NMR Biomed.* 29, 1088–1097. doi: 10.1002/nbm.3567
- Bartus, M., Lomnicka, M., Kostogrys, R., Kaźmierczak, P., Watała, C., Słominska, E. M., et al. (2008). 1-methylnicotinamide (MNA) prevents endothelial dysfunction in hypertriglyceridemic and diabetic rats. *Pharm. Rep.* 60, 127–138.
- Blaziejczyk, A., Switalska, M., Chlopicki, S., Marcinek, A., Gebicki, J., Nowak, M., et al. (2016). 1-Methylnicotinamide and its structural analog 1,4-dimethylpyridine for the prevention of cancer metastasis. *J. Exp. Clin. Cancer Res.* 35:110. doi: 10.1186/s13046-016-0389-9
- Böger, R. H. (2003). The emerging role of asymmetric dimethylarginine as a novel cardiovascular risk factor. *Cardiovasc. Res.* 59, 824–833. doi: 10.1016/S0008-6363(03)00500-5
- Böger, R. H., Bode-Böger, S. M., Sydow, K., Heistad, D. D., and Lentz, S. R. (2000). Plasma concentration of asymmetric dimethylarginine, an endogenous inhibitor of nitric oxide synthase, is elevated in monkeys with hyperhomocyst(e)inemia or hypercholesterolemia. *Arterioscler. Thromb. Vasc. Biol.* 20, 1557–1564. doi: 10.1161/01.ATV.20.6.1557
- Böger, R. H., Bode-Böger, S. M., Szuba, A., Tsao, P. S., Chan, J. R., Tangphao, O., et al. (1998). Asymmetric dimethylarginine (ADMA): a novel risk factor for endothelial dysfunction: its role hypercholesterolemia. *Circulation* 98, 1842–1847. doi: 10.1161/01.CIR.98.18.1842
- Böger, R. H., Bode-Böger, S. M., Thiele, W., Junker, W., Alexander, K., and Frölich, J. C. (1997). Biochemical evidence for impaired nitric oxide synthesis in patients with peripheral arterial occlusive disease. *Circulation* 95, 2068–2074. doi: 10.1161/01.CIR.95.8.2068
- Botnar, R. M., and Makowski, M. R. (2012). Cardiovascular magnetic resonance imaging in small animals. *Prog. Mol. Biol. Transl. Sci.* 105, 227–261. doi: 10.1016/B978-0-12-394596-9.00008-1
- Bryniarski, K., Biedron, R., Jakubowski, A., Chlopicki, S., and Marcinkiewicz, J. (2008). Anti-inflammatory effect of 1-methylnicotinamide in contact hypersensitivity to oxazolone in mice; involvement of prostacyclin. *Eur. J. Pharmacol.* 578, 332–338. doi: 10.1016/j.ejphar.2007.09.011
- Brzozowski, T., Konturek, P. S., Chlopicki, S., Sliwowski, Z., Pawlik, M., Ptak-Belowska, A., et al. (2008). Therapeutic potential of 1-methylnicotinamide against acute gastric lesions induced by stress: role of endogenous prostacyclin and sensory nerves. *J. Pharmacol. Exp. Ther.* 326, 105–116. doi: 10.1124/jpet.108.136457
- Cahill, P. A., and Redmond, E. M. (2016). Vascular endothelium – gatekeeper of vessel health. *Atherosclerosis* 248, 97–109. doi: 10.1016/j.atherosclerosis.2016.03.007
- Candido, R., Jandeleit-Dahm, K. A., Cao, Z., Nesteroff, S. P., Burns, W. C., Twigg, S. M., et al. (2002). Prevention of accelerated atherosclerosis by angiotensin-converting enzyme inhibition in diabetic apolipoprotein E-deficient mice. *Circulation* 106, 246–253. doi: 10.1161/01.CIR.0000021122.63813.32
- Canton, G., Hippe, D. S., Sun, J., Underhill, H. R., Kerwin, W. S., Tang, D., et al. (2012). Characterization of distensibility, plaque burden, and composition of the atherosclerotic carotid artery using magnetic resonance imaging. *Med. Phys.* 39, 6247–6253. doi: 10.1118/1.4754302
- Cavalcante, J. L., Lima, J. A., Redheuil, A., and Al-Mallah, M. H. (2011). Aortic stiffness: current understanding and future directions. *J. Am. Coll. Cardiol.* 57, 1511–1522. doi: 10.1016/j.jacc.2010.12.017
- Cheng, H. L., and Wright, G. A. (2006). Rapid high-resolution T1 mapping by variable flip angles: accurate and precise measurements in the presence of radiofrequency field inhomogeneity. *Magn. Reson. Med.* 55, 566–574. doi: 10.1002/mrm.20791
- Chlopicki, S., and Gryglewski, R. J. (2005). Angiotensin converting enzyme (ACE) and Hydroxymethylglutaryl-CoA (HMG-CoA) reductase inhibitors in the forefront of pharmacology of endothelium. *Pharm. Rep.* 57(Suppl.), 86–96.
- Chlopicki, S., Swies, J., Mogielnicki, A., Buczek, W., Bartus, M., Lomnicka, M., et al. (2007). 1-Methylnicotinamide (MNA), a primary metabolite of nicotinamide, exerts anti-thrombotic activity mediated by a cyclooxygenase-2/prostacyclin pathway. *Br. J. Pharmacol.* 152, 230–239. doi: 10.1038/sj.bjp.0707383
- Closs, E. I., Ostad, M. A., Simon, A., Warnholtz, A., Jabs, A., Habermeier, A., et al. (2012). Impairment of the extrusion transporter for asymmetric dimethyl-L-arginine: a novel mechanism underlying vasospastic angina. *Biochem. Biophys. Res. Commun.* 423, 218–223. doi: 10.1016/j.bbrc.2012.05.044
- Crauwels, H. M., Van Hove, C. E., Holvoet, P., Herman, A. G., and Bult, H. (2003). Plaque-associated endothelial dysfunction in apolipoprotein E-deficient mice on a regular diet. effect of human apolipoprotein AI. *Cardiovasc. Res.* 59, 189–199. doi: 10.1016/S0008-6363(03)00353-5
- Csányi, G., Gajda, M., Franczyk-Zarow, M., Kostogrys, R., Gwoźdź, P., Mateuszuk, L., et al. (2012). Functional alterations in endothelial NO, PGI2 and EDHF pathways in aorta in ApoE/LDLR-/- mice. *Prostaglandins Other Lipid Mediat.* 98, 107–115. doi: 10.1016/j.prostaglandins.2012.02.002
- Dharmashankar, K., and Widlansky, M. E. (2010). Vascular endothelial function and hypertension: insights and directions. *Curr. Hypertens. Rep.* 12, 448–455. doi: 10.1007/s11906-010-0150-2
- Dong, B., Zhang, C., Feng, J. B., Zhao, Y. X., Li, S. Y., Yang, Y. P., et al. (2008). Overexpression of ACE2 enhances plaque stability in a rabbit model of atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 28, 1270–1276. doi: 10.1161/ATVBAHA.108.164715
- Franses, J. W., Drosu, N. C., Gibson, W. J., Chitalia, V. C., and Edelman, E. R. (2013). Dysfunctional endothelial cells directly stimulate cancer inflammation and metastasis. *Int. J. Cancer* 133, 1334–1344. doi: 10.1002/ijc.28146
- Frolow, M., Drozd, A., Kowalewska, A., Nizankowski, R., and Chlopicki, S. (2015). Comprehensive assessment of vascular health in patients; towards endothelium-guided therapy. *Pharm. Rep.* 67, 786–792. doi: 10.1016/j.pharep.2015.05.010
- Gebicki, J., Sysa-Jedrzejowska, A., Adamus, J., Woźniacka, A., Rybak, M., and Zielonka, J. (2003). 1-methylnicotinamide: a potent anti-inflammatory agent of vitamin origin. *Pol. J. Pharm.* 55, 109–112.
- Gotschy, A., Bauer, E., Schrodt, C., Lykowsky, G., Ye, Y. X., Rommel, E., et al. (2013). Local arterial stiffening assessed by MRI precedes atherosclerotic plaque formation. *Circulation* 127, 916–923. doi: 10.1161/CIRCIMAGING.113.000611



- Gryglewski, R. J., Chlopicki, S., and Swies, J. (2005). In vivo endothelial interaction between ACE and COX inhibitors. *Prostaglandins Leukot. Essent. Fatty Acids* 72, 129–131. doi: 10.1016/j.plefa.2004.10.010
- Gryglewski, R. J., Swies, J., Uracz, W., Chlopicki, S., and Marcinkiewicz, E. (2003). Mechanisms of angiotensin-converting enzyme inhibitor induced thrombolysis in wistar rats. *Thromb. Res.* 110, 323–329. doi: 10.1016/j.thromres.2003.08.005
- Halcox, J. P., Schenke, W. H., Zalos, G., Mincemoyer, R., Prasad, A., Waclawiw, M. A., et al. (2002). Prognostic value of coronary vascular endothelial dysfunction. *Circulation* 106, 653–658. doi: 10.1161/01.CIR.0000025404.78001.D8
- Herold, V., Wellen, J., Ziener, C. H., Weber, T., Hiller, K. H., Nordbeck, P., et al. (2009). In vivo comparison of atherosclerotic plaque progression with vessel wall strain and blood flow velocity in apoE(-/-) mice with MR microscopy at 17.6 T. *MAGMA* 22, 159–166. doi: 10.1007/s10334-008-0160-0
- Huang, A., Pinto, J. T., Froogh, G., Kandhi, S., Qin, J., Wolin, M. S., et al. (2015). Role of homocysteinylation of ACE in endothelial dysfunction of arteries. *Am. J. Physiol. Heart Circ. Physiol.* 308, H92–H100. doi: 10.1152/ajpheart.00577.2014
- Jakubowski, A., Sternak, M., Jablonski, K., Cizek-Lenda, M., Marcinkiewicz, J., and Chlopicki, S. (2016). 1-methylnicotinamide protects against liver injury induced by concanavalin A via a prostacyclin-dependent mechanism: a possible involvement of IL-4 and TNF- $\alpha$ . *Int. Immunopharmacol.* 31, 98–104. doi: 10.1016/j.intimp.2015.11.032
- Jiang, F., Yang, J., Zhang, Y., Dong, M., Wang, S., Zhang, Q., et al. (2014). Angiotensin-converting enzyme 2 and angiotensin 1–7: novel therapeutic targets. *Nat. Rev. Cardiol.* 11, 413–426. doi: 10.1038/nrcardio.2014.59
- Jiang, N., Wang, M., Song, J., Liu, Y., Chen, H., Mu, D., et al. (2016). N-methylnicotinamide protects against endothelial dysfunction and attenuates atherogenesis in apolipoprotein E-deficient mice. *Mol. Nutr. Food Res.* 60, 1625–1636. doi: 10.1002/mnfr.201400013.This
- Kao, C. H., Chen, J. K., Kuo, J. S., and Yang, V. C. (1995). Visualization of the transport pathways of low density lipoproteins across the endothelial cells in the branched regions of rat arteries. *Atherosclerosis* 116, 27–41. doi: 10.1016/0021-9150(95)05519-3
- Karbach, S., Wenzel, P., Waisman, A., Munzel, T., and Daiber, A. (2014). eNOS uncoupling in cardiovascular diseases—the role of oxidative stress and inflammation. *Curr. Pharm. Des.* 20, 3579–3594. doi: 10.2174/13816128113196660748
- Kostogrys, R. B., Franczyk-Żarów, M., Maślak, E., Gajda, M., Mateuszuk, L., Jackson, C. L., et al. (2012). Low carbohydrate, high protein diet promotes atherosclerosis in apolipoprotein E/low-density lipoprotein receptor double knockout mice (apoE/LDLR(-/-)). *Atherosclerosis* 223, 327–331. doi: 10.1016/j.atherosclerosis.2012.05.024
- Lee, J. M., Shirodaria, C., Jackson, C. E., Robson, M. D., Antoniadis, C., Francis, J. M., et al. (2007). Multi-modal magnetic resonance imaging quantifies atherosclerosis and vascular dysfunction in patients with type 2 diabetes mellitus. *Diab. Vasc. Dis. Res.* 4, 44–48. doi: 10.3132/dvdr.2007.005
- Lobbis, M. B., Miserus, R. J., Heeneman, S., Passos, V. L., Mutsaers, P. H., Debernardi, N., et al. (2009). Atherosclerosis: contrast-enhanced mr imaging of vessel wall in rabbit model — comparison of gadofosveset and gadopentetate dimeglumine. *Radiology* 250, 682–691. doi: 10.1148/radiol.250.3080875
- Lovren, F., Pan, Y., Quan, A., Teoh, H., Wang, G., Shukla, P. C., et al. (2008). Angiotensin converting enzyme-2 confers endothelial protection and attenuates atherosclerosis. *Am. J. Physiol. Heart Circ. Physiol.* 295, H1377–H1384. doi: 10.1152/ajpheart.00331.2008
- Ludmer, P. L., Selwyn, A. P., Shook, T. L., Wayne, R. R., Mudge, G. H., Alexander, R. W., et al. (1986). Paradoxical vasoconstriction induced by acetylcholine in atherosclerotic coronary arteries. *N. Engl. J. Med.* 315, 1046–1051. doi: 10.1056/NEJM198610233151702
- Marti, C. N., Gheorghide, M., Kalogeropoulos, A. P., Georgiopoulos, V. V., Quyyumi, A. A., and Butler, J. (2012). Endothelial dysfunction, arterial stiffness, and heart failure. *J. Am. Coll. Cardiol.* 60, 1455–1469. doi: 10.1016/j.jacc.2011.11.082
- Mateuszuk, L., Jasztal, A., Maslak, E., Gasior-Glogowska, M., Baranska, M., Sitek, B., et al. (2016). Antiatherosclerotic effects of 1-methylnicotinamide in apolipoprotein E/low-density lipoprotein receptor-deficient mice: a comparison with nicotinic acid. *J. Pharmacol. Exp. Ther.* 356, 514–524. doi: 10.1124/jpet.115.228643
- Mittal, M., Siddiqui, M. R., Tran, K., Reddy, S. P., and Malik, A. B. (2014). Reactive oxygen species in inflammation and tissue injury. *Antioxid. Redox Signal.* 20, 1126–1167. doi: 10.1089/ars.2012.5149
- Olkowicz, M., Chlopicki, S., and Smolenski, R. T. (2015). Perspectives for angiotensin profiling with liquid chromatography/mass spectrometry to evaluate ACE/ACE2 balance in endothelial dysfunction and vascular pathologies. *Pharmacol. Rep.* 67, 778–785. doi: 10.1016/j.pharep.2015.03.017
- Olkowicz, M., Chlopicki, S., and Smolenski, R. T. (2016). A primer to angiotensin peptide isolation, stability, and analysis by nano-liquid chromatography with mass detection, in Thatcher SE. *Methods Mol. Biol.* (in press).
- Pedersen, S. F., Thrysoe, S. A., Paaske, W. P., Thim, T., Falk, E., Ringgaard, S., et al. (2011). CMR assessment of endothelial damage and angiogenesis in porcine coronary arteries using gadofosveset. *J. Cardiovasc. Magn. Reson.* 13:10. doi: 10.1186/1532-429X-13-10
- Phinikaridou, A., Andia, M. E., Protti, A., Indermuehle, A., Shah, A., Smith, A., et al. (2012). Noninvasive magnetic resonance imaging evaluation of endothelial permeability in murine atherosclerosis using an albumin-binding contrast agent. *Circulation* 126, 707–719. doi: 10.1161/CIRCULATIONAHA.112.092098
- Przyborowski, K., Wojewoda, M., Sitek, B., Zakrzewska, A., Kij, A., Wandzel, K., et al. (2015). Effects of 1-methylnicotinamide (MNA) on exercise capacity and endothelial response in diabetic mice. *PLoS ONE* 10:e0130908. doi: 10.1371/journal.pone.0130908
- Redheuil, A. (2014). Cardiovascular aging: insights from local and regional measures of aortic stiffness using magnetic resonance imaging. *Artery Res.* 8, 66–72. doi: 10.1016/j.artres.2014.01.005
- Schächinger, V., Britten, M. B., and Zeiher, A. M. (2000). Prognostic impact of coronary vasodilator dysfunction on adverse long-term outcome of coronary heart disease. *Circulation* 101, 1899–1906. doi: 10.1161/01.CIR.101.16.1899
- Schnabel, R., Blankenberg, S., Lubos, E., Lackner, K. J., Rupprecht, H. J., Espinola-Klein, C., et al. (2005). Asymmetric dimethylarginine and the risk of cardiovascular events and death in patients with coronary artery disease: results from the atherogene study. *Circ. Res.* 97, e53–e59. doi: 10.1161/01.RES.0000181286.44222.61
- Schulz, E., Wenzel, P., Münzel, T., and Daiber, A. (2014). Mitochondrial redox signaling: interaction of mitochondrial reactive oxygen species with other sources of oxidative stress. *Antioxid. Redox Signal.* 20, 308–324. doi: 10.1089/ars.2012.4609
- Smolenska, Z., Smolenski, R. T., and Zdrojewski, Z. (2016). Plasma concentrations of amino acid and nicotinamide metabolites in rheumatoid arthritis – potential biomarkers of disease activity and drug treatment. *Biomarkers* 21, 218–224. doi: 10.3109/1354750X.2015.1130746
- Sternak, M., Khomich, T. I., Jakubowski, A., Szafarz, M., Szczepański, W., Białas, M., et al. (2010). Nicotinamide N-methyltransferase (NNMT) and 1-methylnicotinamide (MNA) in experimental hepatitis induced by concanavalin A in the mouse. *Pharmacol. Rep.* 62, 483–493. doi: 10.1016/S1734-1140(10)70304-2
- Steven, S., Münzel, T., and Daiber, A. (2015). Exploiting the pleiotropic antioxidant effects of established drugs in cardiovascular disease. *Int. J. Mol. Sci.* 16, 18185–18223. doi: 10.3390/ijms160818185
- Suda, O., Tsutsui, M., Morishita, T., Tasaki, H., Ueno, S., Nakata, S., et al. (2004). Asymmetric dimethylarginine produces vascular lesions in endothelial nitric oxide synthase-deficient mice: involvement of renin-angiotensin system and oxidative stress. *Arterioscler. Thromb. Vasc. Biol.* 24, 1682–1688. doi: 10.1161/01.ATV.0000136656.26019.6e
- Surdacki, A., Nowicki, M., Sandmann, J., Tsikas, D., Boeger, R. H., Bode-Boeger, S. M., et al. (1999). Reduced urinary excretion of nitric oxide metabolites and increased plasma levels of asymmetric dimethylarginine in men with essential hypertension. *J. Cardiovasc. Pharmacol.* 33, 652–658. doi: 10.1097/00005344-199904000-00020
- Teixido-Tura, G., Redheuil, A., Rodríguez-Palomares, J., Gutiérrez, L., Sánchez, V., and Forteza, A., et al. (2014). “Aortic biomechanics by magnetic resonance: early markers of aortic disease in marfan syndrome regardless of aortic dilatation?” *Int. J. Cardiol.* 171, 56–61. doi: 10.1016/j.ijcard.2013.11.044
- Thatcher, S. E., Zhang, X., Howatt, D. A., Lu, H., Gurley, S. B., Daugherty, A., et al. (2011). Angiotensin-converting enzyme 2 deficiency in whole body or



- bone marrow-derived cells increases atherosclerosis in low-density lipoprotein receptor-/- mice. *Arterioscler. Thromb. Vasc. Biol.* 31, 758–765. doi: 10.1161/ATVBAHA.110.221614
- Thomas, M. C., Pickering, R. J., Tsorotes, D., Koitka, A., Sheehy, K., Bernardi, S., et al. (2010). Genetic Ace2 deficiency accentuates vascular inflammation and atherosclerosis in the ApoE knockout mouse. *Circ. Res.* 107, 888–897. doi: 10.1161/CIRCRESAHA.110.219279
- Villeneuve, N., Fortuno, A., Sauvage, M., Fournier, N., Breugnot, C., Jacquemin, C., et al. (2003). Persistence of the nitric oxide pathway in the aorta of hypercholesterolemic apolipoprotein-E-deficient mice. *J. Vasc. Res.* 40, 87–96. doi: 10.1159/000070705
- Walczak, M., Suraj, J., Kus, K., Kij, A., Zakrzewska, A., and Chlopicki, S. (2015). Towards a comprehensive endothelial biomarkers profiling and endothelium-guided pharmacotherapy. *Pharmacol. Rep.* 67, 771–777. doi: 10.1016/j.pharep.2015.06.008
- Wang, J., Qiu, M., Kim, H., and Constable, R. T. (2006). T1 measurements incorporating flip angle calibration and correction in vivo. *J. Magn. Reson.* 182, 283–292. doi: 10.1016/j.jmr.2006.07.005
- Wang, Y., Tikellis, C., Thomas, M. C., and Golledge, J. (2013). Angiotensin converting enzyme 2 and atherosclerosis. *Atherosclerosis* 226, 3–8. doi: 10.1016/j.atherosclerosis.2012.08.018
- Watała, C., Kaźmierczak, P., Dobaczewski, M., Przygodzki, T., Bartuś, M., Łomnicka, M., et al. (2009). Anti-diabetic effects of 1-methylnicotinamide (MNA) in streptozocin-induced diabetes in rats. *Pharmacol. Rep.* 61, 86–98. doi: 10.1016/S1734-1140(09)70010-6
- Wozniacka, A., Wieczorkowska, M., Gebicki, J., and Sysa-Jedrzejowska, A. (2005). Topical application of 1-methylnicotinamide in the treatment of rosacea: a pilot study. *Clin. Exp. Dermatol.* 30, 632–635. doi: 10.1111/j.1365-2230.2005.01908.x
- Zhou, R., Yazdi, A. S., Menu, P., and Tschopp, J. (2011). A role for mitochondria in NLRP3 inflammasome activation. *Nature* 469, 221–225. doi: 10.1038/nature09663
- Zhuo, J. L., Mendelsohn, F. A., and Ohishi, M. (2002). Perindopril alters vascular angiotensin-converting enzyme, AT(1) receptor, and nitric oxide synthase expression in patients with coronary heart disease. *Hypertension* 39 (2 Pt 2), 634–638. doi: 10.1161/hy0202.103417

**Conflict of Interest Statement:** SC is a coinventor of the patent on the use of quaternary pyridinium salts as vasoprotective agents.

The other authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Bar, Olkowicz, Tyrankiewicz, Kus, Jasinski, Smolenski, Skorka and Chlopicki. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.